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Results

II. Allelic Ladder Development

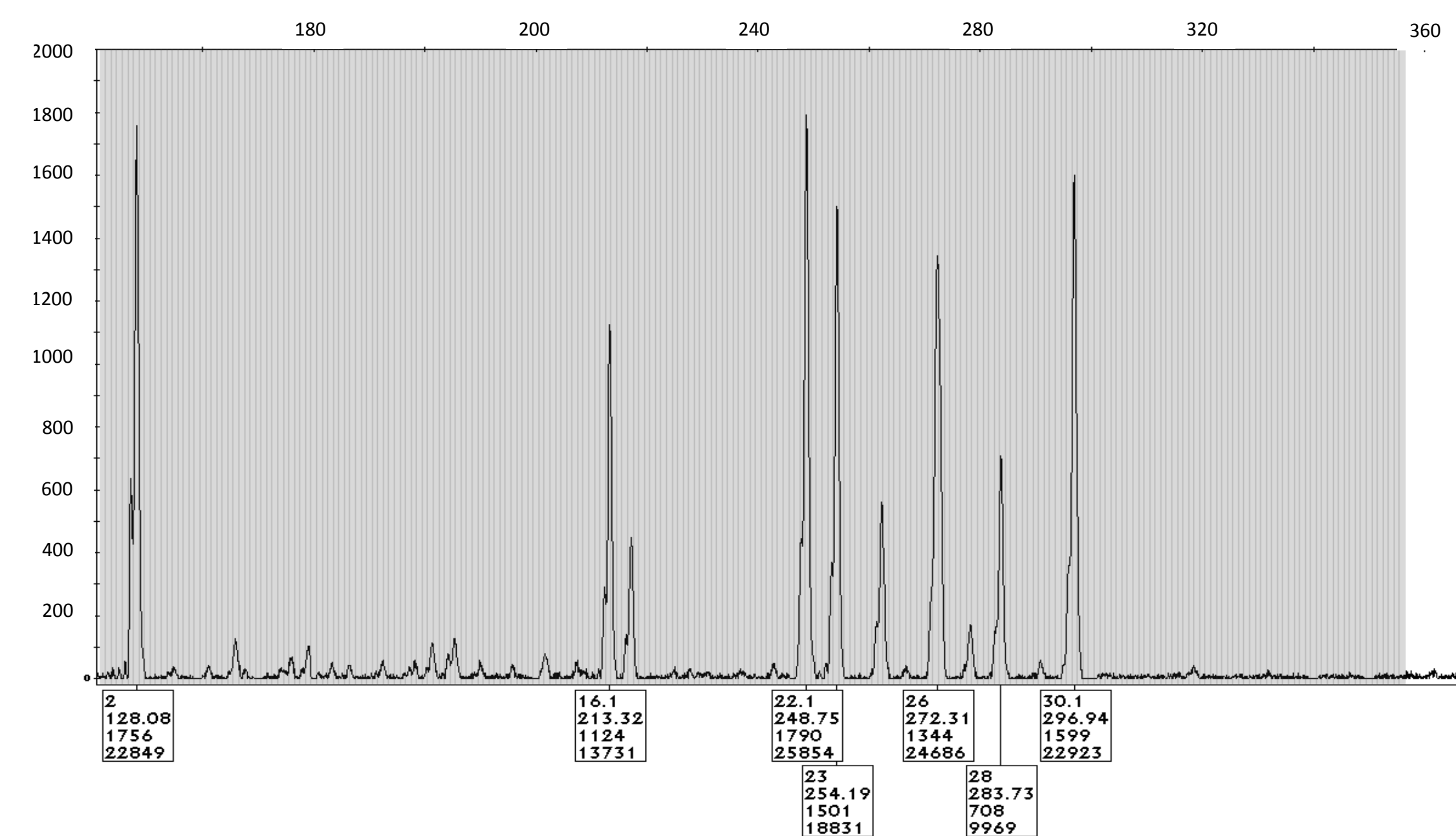
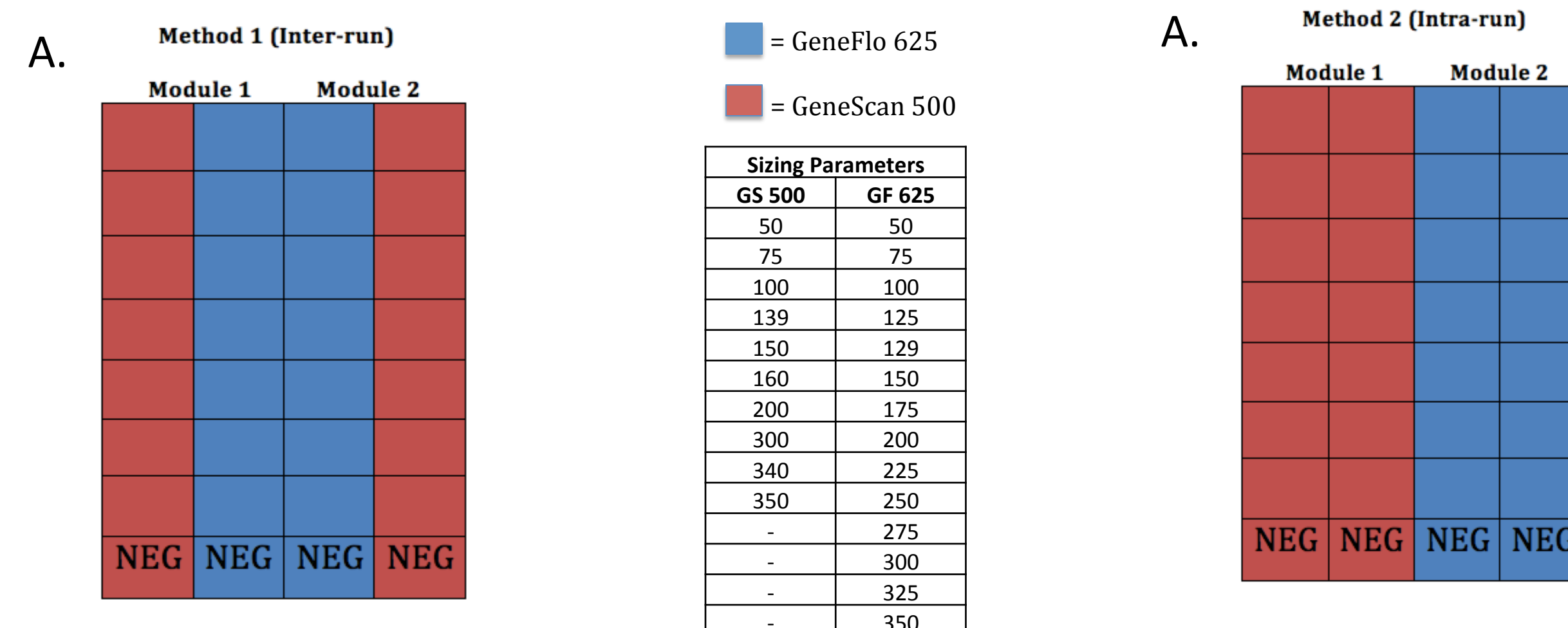


Figure 1. Electropherogram of custom NMI01 allelic ladder

III. Sizing Precision: GeneScan 500 vs. GeneFlo 625

- Legend



- | B. | GS 500 | | | | | | | |
|----|-------------------|----------|----------|----------|----------|----------|----------|--------|
| | Allele 1 | Allele 2 | Allele 3 | Allele 4 | Allele 5 | Allele 6 | Allele 7 | |
| | Average Size (bp) | 128.10 | 213.29 | 248.70 | 254.17 | 272.28 | 283.70 | 296.91 |
| | SD | 0.0949 | 0.1217 | 0.1437 | 0.1521 | 0.1890 | 0.1855 | 0.2269 |
| B. | GS 500 | | | | | | | |
| | Allele 1 | Allele 2 | Allele 3 | Allele 4 | Allele 5 | Allele 6 | Allele 7 | |
| | Average Size (bp) | 127.98 | 213.22 | 248.65 | 254.10 | 272.24 | 283.64 | 296.88 |
| | SD | 0.0855 | 0.1303 | 0.1567 | 0.1638 | 0.1914 | 0.1947 | 0.2274 |
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Table 2. Diagram depicting sample placement for inter-run experiment (A). Data outlining the average fragment size, SD, and SE for each allele in the allelic ladder (B).

Table 3. Diagram depicting sample placement for intra-run experiment (A). Data outlining the average fragment size, SD, and SE for each allele in the allelic ladder (B).

Cannabis treated FTA cards from both 2009 and 2012 all gave NMI01 profiles following extraction, amplification, separation and detection. Even the sample SD, which was a dry plant sample from 2005, was able to obtain a NMI01 profile following FTA card treatment. These results support the concept of using FTA cards as a long-term storage device of *Cannabis* DNA in forensic casework.

Overall, the GeneScan 500 size standard had more precise fragment sizing than the GeneFlo 625 in the inter-run analysis and similar sizing precision in the intra-run analysis. These results agree with past research that has shown the number of fragments in an internal size standard is not a significant factor in sizing precision.⁴ Of particular interest, the GeneScan 500 size standard had consistent standard deviation in fragment sizing between the inter-run and intra-run samples. Despite the high precision (± 0.25 bp), the GeneScan 500 standard displayed an average sizing quality of 0.8714 across the inter- and intra-run analysis versus the GeneFlo 625 standard sizing quality of 0.9937. As alleles ranging from 14 to 30.4 fall within the 200-300 bp region, an allelic ladder is needed to ensure accurate sizing in the current system.

The OSIRIS software successfully analyzed an entire module simulating real-life casework with allelic ladder samples, a positive control, a negative control and suspected *Cannabis* samples. Of particular interest, the samples Box 21 #22 and Box 21 #25, which were obtained from the Mountain and Valley Marijuana Investigation Team (MAVMIT) in an effort to link common origin, gave identical NMI01 profiles (16.1, 22.1) indicating a common origin. Further, when analyzing *Cannabis* samples, the correct allele call was obtained 98.5% of the time with OSIRIS using the custom allelic ladder, while the current system under GeneMapper had 92.4 % accuracy. The OSIRIS software only failed to call the 11.1 allele in sample Box 17 #084 accurately most likely due to the fact that this allele is not present in the custom allelic ladder. Overall, the open source and relatively mobile nature of OSIRIS will potentially further enhance the law enforcement efforts in tracking illegal *Cannabis* activity.

Conclusions

FTA cards treated with *Cannabis* from DEA drug seizures in 2009 and 2012 as well as cards treated with seized *Cannabis* from the UNH Police were chosen for DNA analysis. A 3mm hole punch was taken out of each FTA card before undergoing a washing procedure using TE⁻¹ buffer and FTA Purification Reagent. The resulting FTA cards were amplified using the Terra PCR Direct Genotyping Kit as well as CS1F and CS1R primers.³ The alleles 2, 16.1, 22.1, 23, 26, 28, and 30.1 were chosen to include in the proposed custom NMI01 allelic ladder in this study. Samples from the previous amplifications with these alleles were chosen for the ladder based on amplification efficiency and peak balance. The resulting amplicons were combined in solution following the necessary dilutions to generate a 100 uL stock ladder. The custom allelic ladder was analyzed by both inter- and intra-run fragment analysis in triplicate on an ABI 3130xl Genetic Analyzer. Following this, the sizing software OSIRIS was modified for the NMI01 system and samples were analyzed to simulate real-life case-work.

Due to the effectiveness of STR analysis, there has been a large push to identify additional STR regions in *Cannabis*' DNA in an effort to improve identification accuracy as well as overcome the difficulties present in analyzing *Cannabis* plants generated through clonal propagation.⁵ Overall, the GeneScan 500 size standard appears to size amplicons within the accepted sizing precision, but modifying the system for the automated sizing software OSIRIS along with the use of a further comprehensive allelic ladder may help the current system approach 100% accuracy in correct allele assignment.

Results

I. Analyze NMI01 STR Profiles

A.	Sample ('12)	Allele	Size (bp)	Height (RFU)
	Box 21 #22	16.1	212.94	3350
		22	248.33	2074
	Box 21 #25	16.1	212.74	5138
		22	248.15	2790
	Box 17 #083	22.1	248.52	3555
		23	253.99	4074
	Box 17 #084	11.2	183.83	7446
		26	271.82	2541
	Box 20 #088	23	253.69	2544
		26	271.72	2470
	Box 18 #086	16.1	212.75	1456
		26	271.66	975
	Box 16 #080	16.1	212.72	2124
		25.5	271.41	1374

Table 1. Profiles from Cannabis treated FTA cards from '12 (A), '09 (B), and fresh samples (C) utilizing the GeneScan 5000 for fragment sizing. Highlighted samples indicate those utilized to develop the custom NMI01 allelic ladder.

IV. OSIRIS Sizing Software



Figure 2. Case-work module ran with NMI01 allelic ladder and analyzed via OSIRIS software.

Sample	Box 21 #22		Box 21 #25		6F	4A4	Box 17 #084		Box 20 #088		
Data Points	11		12		12	12	12		12		
File(s)	22.1	16.1	22.1	16.1	26	28	26	11.1	26	23	26
Average Size	212.97	248.99	212.96	248.97	272.36	283.97	271.96	182.57	271.92	254.03	271.84
SD	0.0548	0.0935	0.0835	0.1288	0.0791	0.0612	0.0494	0.0565	0.0724	0.0791	0.084
SE	0.0165	0.0282	0.0241	0.0372	0.0228	0.0177	0.0143	0.0163	0.0209	0.0228	0.024

Table 4. Sizing precision for samples analyzed with OSIRIS using the custom allelic ladder.

Sample	Box 21 #22		Box 21 #25		6F		4A	Box 17 #084		Box 20 #088	
Data Points	12		12		12		12	12		12	
Allele(s)	22.1	16	22.1	16	26	27.5	26	11.2	26	23	26
Average Size	212.71	248.18	212.61	248.06	271.96	283.01	271.65	183.52	271.59	253.59	271.52
SD	0.0762	0.0652	0.0427	0.0567	0.0766	0.0850	0.0748	0.1085	0.0761	0.0792	0.0752
SE	0.0220	0.0188	0.0123	0.0164	0.0221	0.0245	0.0216	0.0313	0.0220	0.0229	0.0191

Table 5. Sizing precision for samples analyzed with GeneMapper.

Sizing Software	Correct Allele Calls	Allele Calls	Allele Call Accuracy (%)	P-value
OSIRIS	128	130	98.5	P<0.001
GeneMapper	122	132	92.5	

Table 6. Allele call accuracy for samples was statistically significant when comparing results between OSIRIS and GeneMapper.

References

1. Allgeier L, Hemenway J, Shirley N, LaNier T, Coyle HM. Field testing of collection cards for *Cannabis sativa* samples with a single hexanucleotide DNA marker. *J Forensic Sci* 2011. 56(5): 1245-1249.
2. John Butler and Bruce McCord workshop at Northeastern Association of Forensic Scientists (Mystic, CT), September 29-30, 2004 "Capillary Electrophoresis in DNA Analysis." http://www.cstl.nist.gov/strbase/pub_pres/NEAFS_DataInterpretation.pdf
3. Hsieh HM, Hou RJ, Tsai LC, Wei CS, Liu SW, Huang LH, Kuo YC, Linacre A, Lee JC. A highly polymorphic STR locus in *Cannabis sativa*. *Forensic Sci Int* 2003. 131(1): 53-58.
4. Klein, S. B., et al. (2003) Addressing ambient temperature variant effects on sizing precision of AmpFISTR Profiler Plus alleles detected on the ABI Prism 310 Genetic Analyzer. *Forensic Science Communications*, 5(1). Available at <https://www.fbi.gov/about-us/lab/forensic-science-communications/fsc/jan2003/index.htm/klein.htm>.
5. Houston R, Birk M, Hughes-Stamm S, Gangitano D. Evaluation of a 13-loci STR multiplex system for *Cannabis sativa* genetic identification. *Int J Legal Med* 2015. DOI 10.1007/s00414-015-1296-x.

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